

Impact of some essential oils on the quality aspect and shelf life of meat

Fahim A. Shaltout¹; Thabet, M.G² and Hanan, A. Koura³

¹ Food Control Department. Faculty of Veterinary Medicine, Benha University ²Animal Health Research Institute (Benha Branch-Food Control Department) ³B.V.Sc., Cairo University, 2000

ABSTRACT

Plant essential oils (EOs) serve as a "safe" alternative to chemical or synthetic antimicrobials and antioxidants to struggle with the food borne pathogens or spoilage organisms, inhibiting lipid oxidation and thus extend shelf life of ground beef application .In our study, we investigated the effectiveness of thyme oil (*thymus vulgaris*) at concentrations (1%, 1.5% and 2%) and cinnamon oil (*cinnamomum zeylanicum*) at concentrations (0.5%, 1% and 1.5%) as natural preservatives as well as their ability to increase the shelf life of minced meat , and also their effects on chemical , microbial analysis and sensory properties of minced meat when been stored at 2°C for 12 days storage period. Obtained results showed that samples treated revealed decreasing of values from chemical, microbial and improving sensory properties than untreated samples (control) ones. Also, thyme oil at concentration 2% and cinnamon oil at concentration 1.5% showed best values than others.

Keywords: essential oils, thyme oil and cinnamon oil, minced meat

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1. INTRODUCTION

Meat has long been known for its nutritive composition which could explain why it is being the first choice sources of animal protein for many people all over the world. Meat has a short shelf life of one day or less at ambient temperature (15-30°C) and a few days at refrigerated temperature (0-10°C) due to microbial spoilage of both pathogenic and non-pathogenic microorganisms and/or lipid oxidation (Houben, 2000). Once the animal has been slaughtered, the meat is fabricated into whole sale or retail cuts. Trim and other cuts of meat are then further processed and ground. This increases the surface area of the meat which allows the increased adherence and growth of the bacteria (Skandamis and Nychas, 2001 and Donsi et al., 2011). Meat has a complex physical structure and chemical composition that is very susceptible to oxidation (Rather et al., 2016). Essential oils are natural, volatile liquid, complex compounds characterized by a strong odor, rarely colored, soluble in lipid and organic solvents.It couldbesynthesizedbyallplantorgans, i.e. buds, f lowers, leaves, stems, twigs, seeds, fruits, root, wood or bark, and are stored in secretary cells, cavities, canals, epidermic cells or glandular trichomes (Bozin et al., 2006)

Thyme (thymus sp.) has much attention due to its high content and wide spectrum of phenolic compounds. antimicrobial and antioxidant properties, and potential for use in meat and meat products (Guttierrez et al., 2008 ; Barbosa et al., 2009 ; Gutierrez et al., 2009 ; Javasena and Jo, 2013 ; Bensid et al., 2014). Thyme essential oil also contains a variety of flavonoids, including a pigenin, naringenin, luteolin and thymonin(Stahl Biskup,1991). The volatile oil components of thyme have also been known to have antimicrobial activity against different bacteria and fungi species (Dorman and Deans 2000).

The bark and leaves of Cinnamon are commonly used as spices in the home kitchen and their distilled essential oils or synthetic analogs are used as flavoring agents in the food and beverage industry (Jham et al., 2005). The major compounds in the essential oil of Cinnamomumzeylanicum are cinnamaldehyde, benzaldehyde, limonene, linalool and eugenol (Baratta et al., 1998). PH values showed the effect of microbial load on minced meat as activation effect of microbial load causes protein hydrolysis with appearance of alkyl group, TBARs values is routinely used as an index of lipid oxidation in minced meat in store and TVN values attributed to the breakdown of proteins as a result of activity of microbial strains and proteolyticenzeymes.

The objective of the present study was the antioxidant as well as the antimicrobial effectiveness of both thyme and cinnamon oils on quality of fresh minced meat during storage at 2°C for 12 days.

2. Materials and methods

Three and half Kg of fresh meat were purchased immediately after slaughter from butcher shops in Kalubiya Governorate and was directly minced and packed in clean polyethylene bags. Then were transported directly in insulated and iced containers to microbiology laboratory (Animal Health Research Institute). Benha, Al-Kalubiya, Egypt.Minced meat sample are divided to two minor groups of treated and untreated (control) ones. The treated groups were divided in to 6 groups (300 g of each) that mixed with thyme at concentrations $((T_1)1\%)$, (T_2) 1.5% and (T_3) 2%) and cinnamon at concentrations $[(T_4)0.5\%, (T_5) 1\%$ and (T_6) 1.5%]. All groups were stored at 2°C and examined periodically every 3 days for chemically, microbiologically and sensory. The experiment was conducted in triplicate for 12 days of storage.

- 1) Chemical analysis:
- a- Measurement of pH according to ISO (1979).

b- Measurement of thiobarbituric acid (TBA) value, the thiobarbituric acid (TBA) assay was carried out according to the procedure of Vyncke (1970).

c- Measurement of total volatile nitrogen (TVN) the total volatile nitrogen (TVN) measurement was carried out according to the procedure ofFAO, (1980).

2) Microbiological analysis:

Preparation of sample, from each group, 10g of meat were taken randomly and aseptically using sterile forceps and scissors, the removed samples were placed in a sterile homogenizer flask contained 45ml of (0.1%) peptone water. The content of each flask were homogenized at 14000 rpm for 2.5 minutes for obtaining a dilution of 10⁻¹, from which 1ml was transferred with a sterile pipette to a sterile test tube containing 9ml of (0.1)peptone water, from which a decimal serial dilution were prepared in a sequential manner up to 10^{-1} , to cover all expected range of samples contamination. For microbial count, colonies were counted and recorded in colony forming units per gram (cfu/g) of meat sampled using the formula: cfu/g= level of

dilution plated \times number of colonies counted / volume plated. These were further expressed in mean colony forming units per gram (mean cfu/g) and converted to \log^{10} base values $(\log^{10}$ cfu/g).

a)Aerobic Plate Counts (APC), according to ICMSF (1996).

b)Enterobacteriaceae Count (EBC),according to ICMSF (1996).

c) Coliform count, according to APHA (1985).

3) Sensory Evaluation:

It was carried out at the third day of the experiment according to Pearson and Tauber (1984). A 9 point hedonic scale (9= Excellent, 8= Very very good, 7= Very good, 6= Good, 5=Medium, 4=Fair, 3=Poor, 2=Very poor, 1=Very very poor) was used for the evaluation of the overall acceptability.

4) Statistical Analysis:

ANOVA was carried out on data of thesensory, chemical and microbiological evaluations.Data are expressed as mean+SE (Gomez and Gomez, 1984).

3.RESULTS

As shown in table (1) pH values at control samples were 6.65, 6.95, 6.99, 7.03 & 7.05, at 0, 3rd,6th,9th & 12th day of storage period, respectively. Samples treated by thyme oil the pH values at concentration 1% (T1) were 6.65, 6.80, 6.77, 6.68 & 6.75; concentration 1.5% (T2) pH values were 6.65, 6.83, 6.81, 6.78 &6.72 and concentration 2% (T3) pH values were 6.65, 6.39, 6.37, 6.31& 6.26, at 0, 3rd day,6th day,9th day&12th day of storage period, respectively. By using cinnamon (C. zeylanicum) oil the pH values at concentration 0.5% (T4) were 6.65, 6.82, 6.79, 6.74 & 6.70; at concentration 1% (T5) values were 6.65, 6.80, 6.77, 6.72 & 6.66 and at concentration 1.5% (T6) pH values were 6.65, 6.33, 6.31, 6.28 & 6.26, at 0, 3rd day, 6th day, 9th day&12th day of storage period, respectively.

As shown in table (2) results showed that TBA values at control samples were 0.140,0.168,0.178, 0.190 & 0.221 at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively. By using thyme oil at concentration 1% (T1) the TBA reactive substance values were 0.140, 0.152, 0.161, 0.172 & 0.177; using concentration 1.5% (T2) TBA values were 0.138, 0.163, 0.176, 0.182 & 0.191 and using concentration 2% (T3) TBA values were 0.137, 0.140, 0.140, 0.142 & 0.143, at 0 day, 3th day, 6th day,9th day & 12th day, respectively. By using cinnamon oil at concentration 0.5% (T4) TBA values were 0.140, 0.152, 0.161, 0.172 & 0.186; using concentration 1% (T5) TBA values were 0.139, 0.163, 0.176, 0.182 & 0.195 and using concentration 1.5% (T6) TBA values were 0.139, 0.140, 0.140, 0.142 & 0.148. at 0 day, 3rd day,6th day,9th day and 12th day of storage period, respectively.

As shown in table (3) at control sample TVN values were 6.25, 39.8, 58.2, 72.44 & 84.21, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively. At samples treated with thyme oil at concentration 1% (T1) TVN values were 6.25, 18.23, 27.1, 35.22 & 44.22.; at concentration 1.5% (T2) TVN values were 6.25, 18.1, 26.4, 31.1&41.1 and at concentration 2%(T3) TVN values were 6.24, 16.2, 25.1, 28.65 & 35.21, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively. By using cinnamon oil at concentration 0.5% (T4) TVN values were 6.25, 18.7, 28.8, 36.1 & 50.12.; at concentration 1% (T5) TVN values were 6.25, 17.99, 26.9, 35.4 &45.14 and at concentration 1.5%(T6) TVN values were 6.25, 16.4, 26.1, 31.3 & 35.42, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively.

As shown in table (4) results showed that the APC values of control minced meat samples were 5.19×10^5 , 6.11×10^7 , 4.15×10^9 , 2.25×10^{10}

 $\&2.33 \times 10^{12}$, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively. By using thyme oil at concentration 1% (T1) APC values were 5.19×10^5 , 7.63×10^6 , 5.15x10⁹& 6.25×10^7 . 5.10×10^{10} .: at concentration 1.5% (T2) APC values were 5.19×10^5 , 4.53×10^6 , 5.11×10^7 , 4.22×10^8 & 3.13x10⁹ and at concentration 2% (T3) APC 5.18×10^5 , 3.73×10^6 , 3.15×10^7 , were $5.89 \times 10^8 \& 2.15 \times 10^8$, at 0 day, 3^{rd} day, 6^{th} day, day&12th day of storage period, 9th respectively. By using cinnamon oil at concentration 0.5%(T4) APC values were 5.19×10^5 , 8.53×10^6 , 6.12×10^7 , 5.15×10^8 & 5.14x10¹⁰; at concentration 1%(T5) APC values were 5.19x10⁵, 7.93x10⁶, 6.10x10⁸, $5.15 \times 10^8 \& 4.32 \times 10^9$ and at concentration 1.5%(T6) APC values were 5.19×10^5 , 5.12×10^6 , 4.14×10^8 , $3.15 \times 10^8 \& 2.16 \times 10^9$, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively.

As shown in table (5) results showed that the enterobacteriacea count EBC at control sample was 9.32×10^3 , 7.24×10^4 , 8.15×10^5 , $4.25 \times 10^6 \& 3.25 \times 10^8$, at 0 day, 3rd day, 6th day, 9thday&12thday of storage period, respectively. By using thyme oil at concentration 1% (T1) EBC was 9.32x10³, 7.04×10^4 , 7.12×10^5 , $6.45 \times 10^5 \& 6.25 \times 10^6$.; at concentration 1.5% (T2) EBC was 9.32x10³, 7.03×10^4 , 4.11×10^5 , $9.25 \times 10^4 \& 4.25 \times 10^5$ and at concentration 2% (T3) EBC was 9.32x10³, 6.81×10^4 , 8.65×10^4 , $8.25 \times 10^3 \& 5.25 \times 10^4$, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively. %). By using cinnamon oil at concentration 0.5% (T4) (EBC) was 9.32×10^3 , 7.24×10^4 , 7.24×10^5 , $5.27 \times 10^5 \& 8.95 \times 10^6$.; at concentration 1% (T5) EBC was 9.32×10^3 , 7.01×10^4 , 7.15×10^5 , $2.25 \times 10^5 \& 3.25 \times 10^6$ • at concentration 1.5%(T6) EBC was 9.32x10³, 6.92x10⁴, 8.95x10⁴, 6.25x10⁴&4.35x10⁵. At 0

day, 3rd day, 6th day, 9thday&12thday of storage period, respectively.

As shown in table (6) results showed that coliform count at control samples was 6.45×10^3 , 4.23×10^5 , 3.45×10^6 , $5.53 \times 10^7 \&$ 6.45×10^8 , at 0 day, 3^{rd} day, 6th day. 9thday&12thday of storage period, respectively. By using thyme oil at concentration1%(T1) coli form count was 6.45×10^3 . 3.21×10^5 . 2.45×10^5 . $6.13 \times 10^5 \& 6.15 \times 10^6$; at concentration 1.5% (T2) coliform count was 6.45×10^3 , 4.53×10^4 , 3.35×10^5 , $3.03 \times 10^5 \& 5.45 \times 10^6$ and coliform count at concentration 2%(T3) was 6.25×10^3 , 3.23×10^4 , 2.15×10^4 , $8.53 \times 10^5 \& 4.45 \times 10^5$, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively. By using cinnamon oil at concentration 0.5% (T4) coliform count was 6.45×10^3 , 4.13×10^4 , 5.25×10^5 , $9.23 \times 10^5 \& 6.25 \times 10^6$.; at concentration 1% (T5) coliform count was 6.45×10^3 . 2.43×10^4 . 3.15×10^5 . $2.44 \times 10^5 \& 6.05 \times 10^6$ and at concentration 1.5% (T6) coliform count was 6.45×10^3 , 4.12×10^4 , 6.45×10^5 , $5.63 \times 10^5 \& 5.55 \times 10^6$, at 0 day, 3rd day, 6th day, 9th day&12th day of storage period, respectively.

As shown in table (7) results shown that sensory properties of different treated minced beef samples during cold storage (2°C) were enhanced by using thyme and cinnamon oils compared to the untreated (control) samples at0 day,3rd day,6th day,9th day and 12th day of the storage period, respectively. Generally, samples containing 2% thyme oil and 1.5% cinnamon oil, demonstrated the highest enhancement of sensory attributes, while the samples treated with 1% of thyme and 0.5% cinnamon oils demonstrated lower enhancement.

Storage days	pH values \pm SE						
	0	3	6	9	12		
Control	6.65 ± 0.02^{a}	6.95±0.01 ^a	6.99 ±0.02 ^{ab}	7.03 ± 0.02 ^c	7.05 ± 0.03^{d}		
Thyme 1%(T1)	6.65 ± 0.02^{a}	6.80 ± 0.01^{a}	6.77 ± 0.01^{ab}	6.68±0.01 ^c	6.75 ± 0.01^{d}		
Thyme 1.5%(T2)	6.65 ± 0.02^{a}	6.83±0.03 ^a	6.81 ± 0.01^{ab}	6.78±0.01 ^c	6.72 ± 0.01^{d}		
Thyme 2%(T3)	6.65 ± 0.02^{a}	6.39±0.04 ^a	6.37±0.01 ^{ab}	6.31±0.01 °	6.26 ± 0.01^{d}		
C. zeylanicum 0.5%(T4)	6.65 ± 0.02^{a}	$6.82\pm\!0.01^{a}$	6.79 ± 0.02^{ab}	6.74 ± 0.01^{b}	6.70±0.01 ^c		
C. zeylanicum 1%(T5)	6.65 ± 0.02^{a}	6.80 ± 0.03^{a}	6.77 ± 0.01^{ab}	6.72 ± 0.01^{b}	$6.66 \pm 0.01^{\circ}$		
C. zeylanicum 1.5%(T6)	6.65 ± 0.02^{a}	6.33 ± 0.02^{a}	6.31 ± 0.01^{ab}	6.28 ± 0.01^{b}	$6.26 \pm 0.01^{\circ}$		

Table	(1)	Mean	PH	values	of the	examined	samples	of mince	ed meat	stored at	2°C	(Mean	+ SE).
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Values with different letters within the same row differed significantly at (P<0.05).

Table (2): Mean values of thiobarbituric acid reactive substances (TBARs) of examined minced meat during cold storage at 2°C (Mean ± SE).

Storage days		TBA mg Malonaldehyde/Kg ±SE							
	0	3	6	9	12				
Control	0.140 ± 0.01^{a}	0.168 ± 0.02^{b}	0.178±0.01 ^c	0.190±0.01 ^c	0.221 ± 0.01^{d}				
T1	0.140 ± 0.01^{a}	0.152±0.01 ^a	0.161 ± 0.01^{b}	$0.172 \pm 0.01^{\circ}$	$0.177 \pm 0.01^{\circ}$				
T2	0.138±0.02 ^a	0.163 ± 0.02^{b}	0.176 ± 0.01^{b}	0.182 ± 0.02^{b}	0.191±0.01 ^c				
T3	0.137±0.01 ^a	0.140±0.01 ^a	0.140 ± 0.00^{a}	0.142 ± 0.01^{b}	0.143 ± 0.02^{b}				
T4	0.140±0.01 ^a	0.152±0.01 ^a	0.161±0.01 ^b	0.172±0.01 ^c	0.186 ± 0.02^{d}				
T5	0.139±0.02 ^a	0.163 ± 0.02^{b}	0.176 ± 0.01^{b}	0.182 ± 0.02^{b}	$0.195 \pm 0.01^{\circ}$				
T6	0.139±0.01 ^a	0.140±0.01 ^a	0.140 ± 0.00^{a}	0.142 ± 0.01^{b}	$0.148 \pm 0.02^{\circ}$				

Values with different letters within the same row differed significantly at (P<0.05).

Table (3): Mean values of total volatile nitrogen (TVN) of examined minced meat stored at 2°C (Mean ± SE).

Storage days	TVN values						
	0	3	6	9	12		
Control	6.25±1.35	39.80±1.7	58.20±1.22	72.44±1.37	84.21±1.31		
Thyme 1% (T1)	6.25±1.34	18.23±1.8	27.1±1.3	35.22±1.3	44.22±1.68		
Thyme 1.5% (T2)	6.25±1.32	18.1±1.55	26.4±1.32	31.1±1.25	41.10±1.70		
Thyme 2% (T3)	6.24±1.25	16.2±1.33	25.1±1.22	28.65±1.15	35.21±1.30		
C. zeylanicum0.5% (T4)	6.25±1.35	18.70±1.5	28.8±1.12	36.1±1.15	50.12±1.31		
C. zeylanicum 1% (T5)	6.25±1.35	17.99±1.1	26.9±1.2	35.4±1.45	45.14±1.45		
C. zeylanicum 1.5% (T6)	6.25±1.35	16.4±1.45	26.1±1.4	31.3±1.22	35.42±1.15		

TVN 20 mg / 100 gm raw minced beef (EOS, 2005) + (P< 0.05).

Table (4): Changes in aerobic plate count (APC) of examined minced meat during cold storage at 2°C

Storage days	(APC) log cfu /g \pm SE							
	0	3	6	9	12			
Control	5.19X10 ⁵	$6.11 \text{X} 10^7$	4.15X10 ⁹	$2.25 X 10^{10}$	$2.33 X 10^{12}$			
Thyme 1%(T1)	$5.19X10^{5}$	$7.63 \text{X} 10^{6}$	$6.25 \text{X} 10^7$	5.15X10 ⁹	$5.10 \text{X} 10^{10}$			
Thyme 1.5% (T2)	5.19×10^{5}	$4.53 X 10^{6}$	$5.11X10^{7}$	$4.22X10^{8}$	3.13X10 ⁹			
Thyme 2% (T3)	5.18×10^{5}	3.73×10^{6}	$3.15 \text{X} 10^7$	$2.89X10^{8}$	2.15×10^{8}			
C. zeylanicum 0.5% (T4)	5.19×10^{5}	$8.53 X 10^{6}$	$6.12 \text{X} 10^7$	5.15X10 ⁸	$5.14 \text{X} 10^{10}$			
C. zeylanicum 1%(T5)	5.19×10^{5}	7.93×10^{6}	$6.10 \mathrm{X10^8}$	5.15X10 ⁸	4.32×10^{9}			
C. zeylanicum 1.5%(T5)	$5.19 \text{X} 10^5$	$5.21X10^{6}$	$4.14 \mathrm{X10^8}$	3.15X10 ⁸	2.16×10^{9}			

Values with different letters within the same row differed significantly at (P<0.05).

Table (5): Changes in	enterobacteriaceae cour	t of examined mir	nced meat during co	ld storage at 2°C (M	1ean ±
SE).					

Storage days	Enterobacteriaceae count							
	0	3	6	9	12			
Control	9.32x10 ³	7.24x10 ⁴	8.15x10 ⁵	4.25x10 ⁶	3.25x10 ⁸			
	$\pm 2.25 \times 10^3$	$\pm 1.1 \times 10^4$	±1.89x10 ⁵	$\pm 2.09 \times 10^{6}$	$\pm 2.19 \times 10^{8}$			
Thyme 1% (T1)	9.32x10 ³	7.04x10 ⁴	7.12x10 ⁵	6.45x10 ⁵	6.25x10 ⁶			
	$\pm 2.25 \times 10^{3}$	$\pm 1.1 \times 10^4$	$\pm 1.17 \times 10^{5}$	$\pm 1.09 \times 10^{5}$	$\pm 2.19 \times 10^{6}$			
Thyme 1.5%(T2)	9.32x10	7.03x10 ⁴	4.11x10 ⁵	9.25x10 ⁴	4.25x10 ⁵			
-	$^{3}\pm2.25x10^{3}$	$\pm 1.1 \times 10^4$	$\pm 1.99 \times 10^5$	$\pm 2.09 \text{x} 10^4$	$\pm 1.19 \times 10^{5}$			
Thyme 2%(T3)	9.32x10 ³	6.81x10 ⁴	8.65x10 ⁴	8.25x10 ³	5.25x10 ⁴			
-	$\pm 2.25 \times 10^{3}$	$\pm 1.1 \times 10^4$	$\pm 2.36 \times 10^4$	$\pm 2.69 \times 10^3$	$\pm 2.19 \times 10^4$			
C. zeylanicum 0.5% (T4)	9.32x10 ³	7.24x10 ⁴	7.24x10 ⁵	5.27x10 ⁵	8.95x10 ⁶			
	$\pm 2.25 \times 10^{3}$	$\pm 1.1 \times 10^4$	$\pm 1.81 \times 10^5$	$\pm 1.67 \times 10^5$	$\pm 2.19 \times 10^{6}$			
C. zeylanicum 1 % (T5)	9.32×10^3	7.01x10 ⁴	7.15x10 ⁵	2.25x10 ⁵	3.25x10 ⁶			
	$\pm 2.25 \times 10^3$	$\pm 1.1 \times 10^4$	$\pm 1.29 \times 10^5$	$\pm 2.09 \times 10^5$	$\pm 2.01 \times 10^{6}$			
C. zeylanicum 1.5% (T6)	9.32x10 ³	6.92x10 ⁴	8.95x10 ⁴	6.25x10 ⁴	4.35x10 ⁵			
	$\pm 2.25 \times 10^{3}$	$\pm 1.1 \times 10^4$	$\pm 2.34 \times 10^4$	$\pm 2.43 \times 10^4$	$\pm 1.17 \times 10^5$			

Table(6): Changes in coli form count of examined minced meat during cold storage at $2^{\circ}C$ (Mean ±SE).

Storage days	0	3	6	9	12
Control	6.45×10^3	4.23×10^5	3.45×10^{6}	5.53×10^7	$6.45 \text{x} 10^8$
	$\pm 1.21 \times 10^{3}$	$\pm 1.01 \mathrm{x} 10^5$	$\pm 1.46 \mathrm{x} 10^{6}$	$\pm 1.11 \text{x} 10^7$	$\pm 1.19 \mathrm{x} 10^{8}$
Thyme 1% (T1)	6.45×10^3	3.21×10^5	2.45×10^5	6.13x10 ⁵	6.15×10^{6}
	$\pm 1.21 \times 10^{3}$	$\pm 1.01 \mathrm{x} 10^5$	$\pm 1.26 \times 10^{5}$	$\pm 1.11 \text{x} 10^5$	$\pm 1.19 \mathrm{x} 10^{6}$
Thyme 1.5% (T2)	6.45×10^3	4.53×10^4	3.35×10^{5}	3.03×10^5	5.45×10^{6}
	$\pm 1.21 \times 10^{3}$	$\pm 1.51 \text{x} 10^4$	$\pm 1.16 \mathrm{x} 10^5$	$\pm 1.11 \text{x} 10^5$	$\pm 1.19 \mathrm{x} 10^{6}$
Thyme 2% (T3)	6.25×10^3	3.23×10^4	2.15×10^4	8.53x10 ⁵	$4.45 \text{x} 10^5$
	$\pm 1.21 \times 10^{3}$	$\pm 1.03 \text{x} 10^4$	$\pm 1.06 \text{x} 10^4$	$\pm 1.11 \text{x} 10^5$	$\pm 1.18 \mathrm{x} 10^5$
C. zeylanicum 0.5% (T4)	6.45×10^3	4.13×10^4	5.25×10^{5}	9.23×10^{5}	6.25×10^{6}
	$\pm 1.21 \times 10^{3}$	$\pm 1.02 x 10^4$	$\pm 1.22 \mathrm{x} 10^5$	$\pm 1.11 x 10^5$	$\pm 1.19 \mathrm{x} 10^{6}$
C. zeylanicum 1 % (T5)	6.45×10^3	2.43×10^4	3.15×10^5	2.44×10^5	$6.05 ext{x} 10^{6}$
	$\pm 1.21 \times 10^{3}$	$\pm 1.01 \mathrm{x} 10^4$	$\pm 1.86 \mathrm{x} 10^5$	$\pm 1.11 x 10^{5}$	$\pm 1.19 \mathrm{x} 10^{6}$
C. zeylanicum 1.5% (T6)	6.45×10^3	4.12×10^4	6.45×10^5	5.63×10^5	$5.55 \text{x} 10^6$
	$\pm 1.21 \times 10^{3}$	$\pm 2.01 \text{x} 10^4$	$\pm 1.26 \times 10^{5}$	$\pm 1.21 \times 10^{5}$	$\pm 1.16 \mathrm{x} 10^{6}$

Table (7). Mean score	s of sensory	v characteristics of	examined	minced me	at during co	old storage	period at '	2°C
14010 (/)	5 01 5 0 11501 j	•	••••••••••				p • • • • • • • • •	

Sensory attributes	Sensory scores						
	0	3 rd	6 th	9 th	12^{th}		
control	Excellent	Fair	poor	Very poor	Very very poor		
T1	Excellent	Good	medium	fair	poor		
T2	Excellent	Good	medium	fair	poor		
Т3	Excellent	Very very good	Very good	good	medium		
T4	Excellent	Good	medium	fair	poor		
T5	Excellent	Very good	good	Medium	fair		
T6	Excellent	Very very good	Very good	good	medium		

4. DISCUSSION

From results in table (1) which showed that samples containing thyme essential oil were lowering in pH values than control samples during different periods of storage test . Also, by increasing concentration of thyme to 2% scored highest effect in lowering pH value than lower concentration (1%). These results were nearly similar to results obtained by Salem et al. (2010) who reported that minced beef mixed by thyme essential oil during cold storage at 4°C withdifferent concentrations of thyme (1.5%, 1%) and 0.5%) showed significant effect in lowering of pH values than those of untreated samples, also the highest oil's concentration was more effective values in lowering PH than lower concentrations were used. These agreement with, Kassemet al. (2011) they stated that pH values of burger samples treated with high thyme concentration of (1.2%)were significantly low (p<0.05) comparing with control samples after4 days of storage at refrigeration temperature, might be due to the antimicrobial effect of phenolic compound in herb oil. By using cinnamon oil, the obtained results indicated that PH values at different concentrations of cinnamon showed lowering pH values than control samples, also by increasing of cinnamon(C.zeylanicum) oil concentrations (1.5%) results showed more lowering in values than lower cinnamon concentrations (0.5%), which may be due to antimicrobial load .These agreement with Yassin - Nessrien, (2003); Gahruie et al.(2017) and Kesavan et al.(2014) who stated that pH values of raw beef meat samples were measured during storage at 4±1°C for 15 days had higher pH value at control samples than samples treated by cinnamon oil, this increase in pH reflects the degree of meat spoilage through protein breakdown for the production of free amino acids, leading to theformation of NH3 and amines, compounds

of alkaline reactions. Also, these results agreement with that obtained by Brilliana et al. (2017) they stated that the addition of cinnamon essential oil on ground beef can lowering pH values of fresh ground beef during refrigerated storage compared to the controlsample, these results because of the active components of the cinnamon essential oil that are cinnamaldehyde, linalool and eugenol.

From results in table (2) which showed that treated samples with thyme oil at different concentrations showed lowering in TBARs values than control samples and increasing of thyme essential oil concentration caused decreasing the TBARs values than lower thyme concentration used. Such findings may be attributed to the high antioxidant effect of thyme essential oil, which is related to the scavenger nature of its flavonoids and phenolic content as apigenin, naringenin, luteolin, thymonin, carvacrol and thymol (Senatore, 1996). Also these results were similar to that was obtained by Salem et al(2010); Kassem et al (2011) and Zengin et al .(2015) who mentioned that TBA values of minced meat were obtained over 9 days of storage at 4°Cand TBA values of control showed a rapid increase with increasing storage time, TBA values of thyme treatment samples on day 0 were significantly lower (P < 0.05) than those for the control sample. Moreover, treatment samples had significantly lower (P < 0.05) TBA values than control at each day of testing throughout sixth and ninth days of the storage period. Also, Ghaderi et al. (2016) who mentioned that changes in TBARS value of beef burgers with different formulations during 8 days of refrigerated storage the amount of TBARs increased suddenly over time in control burgers, and significant differences were evident among treated samples with thyme oil as compared to control samples. By usind cinnamon oil, all treated samples bv cinnamon oil showed lowering in TBARs values than control sample, also increasing concentration of C.zeylanicum essential oil caused decreasing in the TBARs values than lower concentration of the same oil. This result agrees with results obtained by Pikul et al., (1984);Ciftci et al., (2010); Ojagh et al., (2010) and Andevari and Rezaei(2011). These results agree with Kesavan et al. (2014) who recorded that the effect of cinnamon oil on TBARS values of raw beef samples during storage at 4°C for15 days. TBARS values of all treated samples were considerablylower (P < 0.05) than those of control samples during storage, indicating high protection by the spice extracts against lipid oxidation in raw beef meat samples. Also, Irfiana etal. (2017) mentioned that cinnamon oil showed decreasing at TBA values of vacuum-packed ground beef during refrigerated storage (at 0, 4, 8, 12, and 16 day) when compared with TBA values of control samples, as cinnamon oil contains active compounds such as polyphenols, cinnamaldehyde leading to antibacterial action and antioxidant of meat.

From results in table (3) which showed that from the obtained result which appeared that the treatment samples with thyme oil at different concentrations caused decreasing in TVN values than results showed by control samples, also by increasing concentration of thyme (2%) was more effective in decreasing TVN values than lower concentration of thyme (1%), the degree of autolysis and bacterial proteolysisin meat can be measured as TV which actually determines the quantity of amino acids, i.e., tyrosine and tryptophan present in an extract of meat. Nearly agree the results which were obtained by with Salem et al.(2010) who mentioned that preserved minced beef at 4°C during 6 days of storage period recorded the highest rate of TVN values at control samples, also the treated sample with 1.5% thyme oil was more effective in decreasing TVN values than

sample treated with 0.5% thyme oil, due to the role of thyme oil on microbial population and bacterial growth as antimicrobial agents. By using cinnamon oil, from the obtained result which appeared that the treatment samples with cinnamon oil at different concentrations caused decreasing in TVN values than control samples, also by increase concentration of cinnamon (1.5%) was more effective in decreasing TVN values than lower concentration of cinnamon (0.5%)which was used. These results were agree with results were recorded by Quattara et al.(2000) and Biswas et al.(2011)who recorded that lowering TVN values of meat when treated by cinnamon oil than control samples was due to cinnamaldehyde.

From results in table (4) which showed that from the obtained results, samples treated by different concentrations of thyme showed decreasing count of aerobic plate microorganism when compared to control samples, also high concentration of thyme more effective in decreasing this count than lower concentration. These results were similar to results were obtained by Salem et al.(2010); Elgaafary et al.(2016) and Amani (2017) who mentioned that the antibacterial effect of thyme oil in refrigerated minced beef as decreasing APC than control sample. High concentration of thyme oil (1.5%) was more effective on decreasing APC values than values of lower concentration of the same oil By using cinnamon oil, from the (0.5%).obtained results, samples treated with different concentrations of cinnamon caused aerobic decreasing count of plate microorganism when compared to control samples, also high concentration of cinnamon (1.5%) was more effective in decreasing this count than lower concentration of cinnamon oil (0.5%). These results were agree with results were obtained by Krishnamoorthy et al.(2004)they stated that cinnamon (C.zeylanicum) oilconsists of two chemical

volatile phenols and poly phenols. Cinnamon showed as antibacterial properties against gram positive and gram negative bacteria. Also, agree with Irfiana et al. (2017) who recorded that cinnamon oil showed decreasing count of total plat count microorganisms of vacuum-packed ground beef during refrigerated storage (at 0, 4, 8, 12, and 16 day) when compared with count of control samples, due to cinnamon contains active compounds such as polyphenols, cinnamaldehyde and antibacterial compounds.

Results in table (5) showed that samples treated by different concentrations of thyme oil showed decreasing count of enterobacteriaceae when compared to control samples, also high concentration of thyme (2%) was more effective in decreasing this count than lower concentration(1%). These results nearly similar to results were obtained by Bozen et al. (2006) ; Yassin- Nessrien and Abou-Taleb(2007). Also, Salem et al.(2010) stated that preserved minced beef at 4°Calonge 6 days storage period at the control had the highest samples counts of enterobacteriacea compared to other treatment of minced beef by different concentration of thyme (0.5%, 1%, 1.5%). Also, the highest concentration of thyme (1.5%) appeared more decreasing of enterobacteriacea count than lower concentration of thyme (0.5%). Also, Amani (2017) who recorded that preserved minced beef at refrigeration temperature, with treatment by thyme oil play an important role in decreasing enterobacteriacea count especially at 1.5% concentration.

By using cinnamon oil, from the obtained samples results. treated bv different concentrations of cinnamon oil caused decreasing count of enterobacteriacea when compared to control samples, also high concentration of cinnamon was more decreasing of (EBC) lower than concentration of cinnamon (0.5%). These results were nearly similar to results were obtained by Quattara et al. (2000) who mentioned that treatment cooked meat products by trans- cinnamaldehyde, which is main component of cinnamon oil ,produced significant extension of cooked meat products shelf life by inhibiting growth of enterobacteriacea.

Table (6) showed that from the obtained samples results. treated bv different concentrations of thyme oil showed decreasing count of coliform when compared to control samples, also high concentration of thyme oil (2%) was more effective in decreasing this count than lower concentration (1%). These results were nearly similar to that obtained by Hoque et Solomakos al.(2008) et al.(2008); Krischetal. (2010) and Boskovic et al.(2015) Also, Emiroglu et al. (2010) who mentioned that thymol is effective to destroy coliforms when used at concentration of 250,500 and 750 mg/kg of beef minced patties and stored for 16 days under ordinary or modified packaging at refrigeration temperature, also mentioned that covering surface of meat products by 0.8 % thymol, thecount of coliforms decreased by 2-3 log units. Also Salem et al. (2010) stated that preserved minced beef at 4°C alonge 6 days storage period, the control sample had the highest counts of coli forms compared to other treatment of minced beef by different concentration of thyme (0.5%, 1%, 1.5%), highest concentration of thyme (1.5%)appeared more effectiveness in decreasing coliforms count especially at 3rd and 6th day of storage period than lower concentration (0.5%). Also, Amani (2017) who recorded that preserved minced beef at refrigeration temperature, with treatment by thyme oil play an important role in decreasing coliforms count especially at 1.5% concentration. By using cinnamon oil, from the obtained results, samples treated by different concentrations of cinnamon oil caused decreasing count of coliform when compared to control sample, also high concentration of cinnamon (1.5%) was more effective in decreasing this count than lower concentration (0.5%). These results were nearly similar to results were obtained by Hoque et al.(2008); Friedman et al. (2002);Hoque et al.(2007); Charu Gupta et al .(2008) and Reham, (2013) who stated that treatment of minced beef under cool temperature 4°C by using of cinnamon (C.zeylanicum) oil at different concentrations (0.5%, 1%, 1.5%)showed decreasing count coliform when compared to control of sample, also high concentration of cinnamon (1.5%) more effective in decreasing this count concentration (0.5%). than lower Also. Jagadeesh et al.(2013) who mentioned that cinnamon oil decreased coli form count, might be due to cinnamaldehyde and the antimicrobial compound of cinnamon.

Results in table (7) revealed that sensory properties of different treated minced beef samples during cold storage (2°C) were enhanced by using thyme and cinnamon oils compared to the untreated (control) samples at0 day,3rd day,6th day,9th day and 12th day of the storage period, respectively. Generally, samples containing 2% thyme oil and 1.5% cinnamon oil, demonstrated the highest enhancement of sensoryattributes, while the samples treated with 1% of thyme and 0.5% of cinnamon oils demonstrated lower enhancement. This result agrees with that obtained by Sasse et al. (2009) who reported that many herbs and spices as thyme contain antioxidant components that improve both color and flavor stability in meat. Also, Salem et al. (2010) mentioned that sensory properties of minced beef samples during cold storage (4°C) were enhanced by treatment minced beef by different concentrations of thyme oil(0.5%, 1%), 1.5%) compared to the untreated (control) samples, and sample contain 1.5% thyme oil revealed best enhancement of sensory

properties than sample contain 0.5% of the same oil ,at zero, 3rd, and 6th day of the storage period, respectively. Also Ibrahim et al. (2016) mentioned that the sensory properties of different treated minced beef samples during cold storage (4°C) were improved by using different concentrations (.5%, 1% and 1.5%) of thyme oil compared to the control samples after 3hrs, 1st, 2nd, 3rd, 4th and 5th day of the storage period, also sample containing 1.5% thyme demonstrated the highest enhancement of sensory attributes than sample treated with 0.5% of the same oil. Also Amani etal. (2017)they reported that the sensory properties of different treated minced beef samples by using thyme oil at concentrations (0.5% , 1%, 1.5%) and cinnamon oil at concentrations (0.5%, 1%, 1.5%) during cold storage (4°C) were enhanced at treatment samples compared to the untreated (control) samples at zero, 1st, 2^{nd} , 3^{rd} and 4^{th} day of the storage period, also samples containing 1.5% (cinnamon and thyme) oils demonstrated the highest enhancement of sensory properties.

5- Conclusion

Thyme and cinnamon oils can be used as natural meat preservatives with antimicrobial activities against food borne pathogens, and therefore may be useful in maintaining the meat quality, extending shelf- life of meat, preventing economic loss and providing the consumer with food containing natural additives especially at thyme (2%) and cinnamon 1.5%.

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